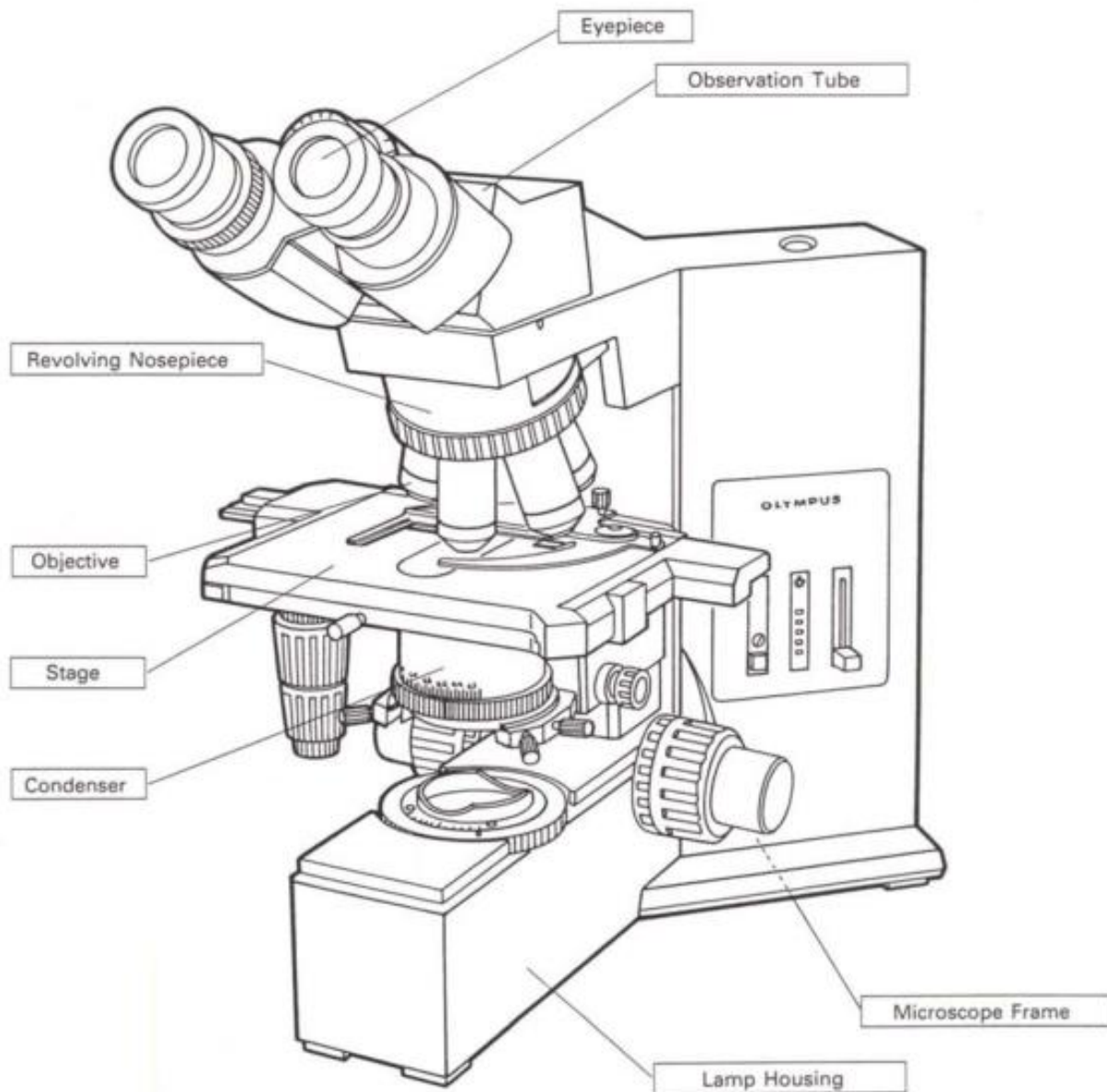


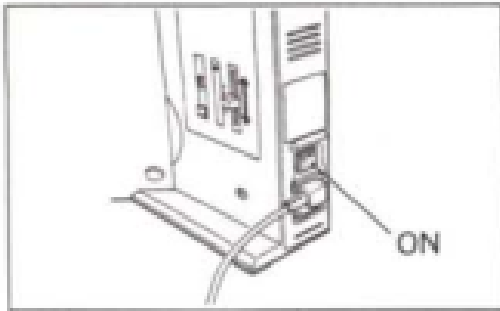
# USER MANUAL BX40 WIDE FIELD MICROSCOPE



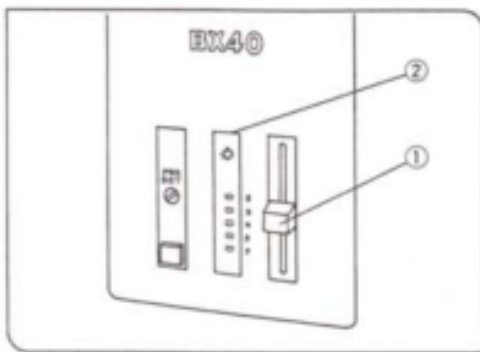
François MICHEL PhD

## STARTING THE SYSTEM

**For white transmission** illumination: switch the power on in the back of the microscope right part



And adjust the brightness with the light intensity lever (1)

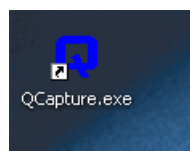


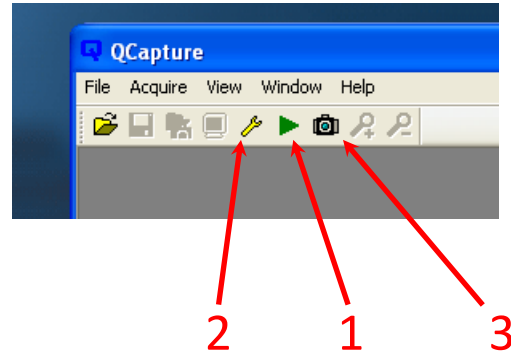
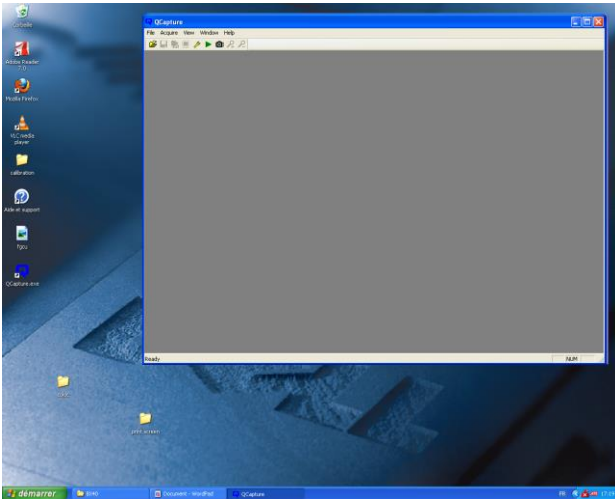
**For fluorescence acquisition** or visualization: switch on the fluorescent lamp  
There are 4 fluorescent cubes: DAPI, GFP, RFP, and deep red (Cy5), their technical characteristics are annexed at the end (p4).

Turn on the computer and log on your **personal session**.

Check if the camera is on (switch and green LED on the top)

Start **Qcapture** software





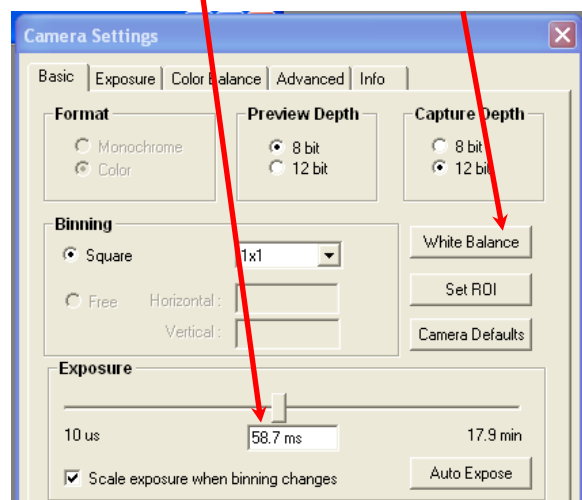
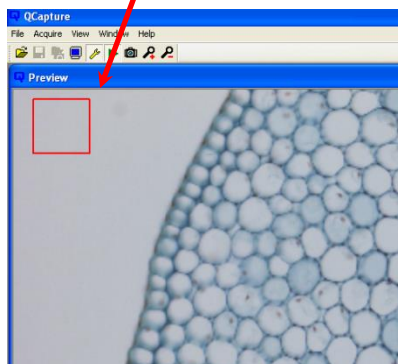
1) Preview (live)

2) Camera Toolbox :

In this window you will: Set the **exposure time** (exposure),

Define white balance for **transmission illumination**:

Create a ROI (drag a rectangle) outside the sample and click on White Balance button



3) Snap a photo (will create another window)

Save it in Tiff format in intranet or external disc.

## TURNING OFF THE SYSTEM

1. Check that all your data are saved
2. Close Qcapture software
3. Logout Windows (Start → Log Off).
4. Switch off the fluo lamp and microscope power

## ANNEXE

### Fluorescent cubes

<u>Fluorophores Cube#</u>	<u>Excitation filter</u>	<u>Dichroic</u>	<u>Emission filter</u>
DAPI 2	363-380	410	435-490
GFP 3	466-498	505	517-555
RFP 4	510-565	560	603-666
Cy5 5	588-650	645	666-740

### Objectives (All Dry) :

<u>Enlargement</u>	<u>N.A.</u>	<u>XY resolution (µm) @ 520nm</u>	<u>Z resolution (µm) @ 520nm</u>	<u>Working distance (mm)</u>
4X	0,10	3,17	104	18,5
10X	0,25	1,27	16,64	10,6
20X	0,5	0,63	4,16	2,2
40X	0,65	0,49	2,46	0,6
60X	0,8	0,40	1,63	0,35

### Measured calibration (1600X1200 pixels)

	<u>pix/µm</u>	<u>µm/pix</u>	<u>Frame size (µm)</u>
4X	0,544	1,84	2941 X 2206
10X	1,36	0,736	1177 X 883
20X	2,71	0,369	590 X 443
40X	5,474	0,182	292 X 219
60X	8,120	0,123	197 X 148