

Depolarizing Actions of GABA in Immature Neurons Depend Neither on Ketone Bodies Nor on Pyruvate

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GABA depolarizes immature neurons because of a high $[Cl^-]_i$ and orchestrates giant depolarizing potential (GDP) generation. Zilberter and coworkers (Rheims et al., 2009; Holmgren et al., 2010) showed recently that the ketone body metabolite DL-3-hydroxybutyrate (DL-BHB) (4 mM), lactate (4 mM), or pyruvate (5 mM) shifted GABA actions to hyperpolarizing, suggesting that the depolarizing effects of GABA are attributable to inadequate energy supply when glucose is the sole energy source. We now report that, in rat pups (postnatal days 4–7), plasma D-BHB, lactate, and pyruvate levels are 0.9, 1.5, and 0.12 mM, respectively. Then, we show that DL-BHB (4 mM) and pyruvate (200 μ M) do not affect (i) the driving force for GABA_A receptor-mediated currents (DF_{GABA}) in cell-attached single-channel recordings, (2) the resting membrane potential and reversal potential of synaptic GABA_A receptor-mediated responses in perforated patch recordings, (3) the action potentials triggered by focal GABA applications, or (4) the GDPs determined with electrophysiological recordings and dynamic two-photon calcium imaging. Only very high nonphysiological concentrations of pyruvate (5 mM) reduced DF_{GABA} and blocked GDPs. Therefore, DL-BHB does not alter GABA signals even at the high concentrations used by Zilberter and colleagues, whereas pyruvate requires exceedingly high nonphysiological concentrations to exert an effect. There is no need to alter conventional glucose enriched artificial CSF to investigate GABA signals in the developing brain.

Introduction

GABA depolarizes and excites immature neurons in many animal species because of their higher $[Cl^-]_i$ compared with mature neurons. This developmental change reflects the sequential operation of the chloride cotransporters NKCC1 and KCC2 (Ben-Ari et al., 1989; Owens et al., 1996; Rivera et al., 1999; Ganguly et al., 2001; Akerman and Cline, 2006) (for review, see Ben-Ari, 2002; Owens and Kriegstein, 2002; Ben-Ari et al., 2007). GABAergic synapses are expressed before glutamatergic synapses, and GABA provides most of the early activity and orchestrates the generation of the first synaptic network-driven giant depolarizing potentials (GDPs) (Ben-Ari et al., 1989; Garaschuk et al., 1998;

Tyzio et al., 1999; Ben-Ari, 2001; Sipila et al., 2006; Crépel et al., 2007; Bonifazi et al., 2009).

Rodent maternal milk is enriched in fatty acids that are transformed in the liver to aceto-acetate and DL-3-hydroxybutyrate (DL-BHB). On the basis of this information, Zilberter and colleagues have recently challenged the developmental sequence of GABA action (Rheims et al., 2009) and reported that adding DL-BHB (4 mM) to artificial CSF (ACSF) almost completely eliminated depolarizing and excitatory actions of GABA. This was suggested to reflect the actions of the Cl^-/HCO_3^- exchanger and not KCC2, reported to be inactive in the neocortex at the ages investigated [postnatal day 1 (P1) to P8] (Rheims et al., 2009). In a subsequent paper, the same group (Holmgren et al., 2010) extended these observations to hippocampal CA3 pyramidal neurons and reported that, in addition to DL-BHB, lactate or pyruvate at exceedingly high concentrations (5 mM) also shifted reversal potential of synaptic GABA_A receptor (GABA_AR)-mediated responses (E_{GABA}) and blocked GDPs, suggesting that the depolarizing actions of GABA are attributable to energy deprivation when glucose is the sole energetic source. Because this suggestion has important implications for neonatal slice studies, we have now reexamined the effects of DL-BHB and pyruvate on GABA actions on neonatal deep layers neocortical neurons and CA3 pyramidal neurons.

Received June 27, 2010; revised Sept. 14, 2010; accepted Sept. 20, 2010.

This work was supported by Inserm, L'Agence Nationale de la Recherche (I.K.), Fédération pour la Recherche sur le Cerveau, European Union Network of European Museum Organisations, Fondation pour la Recherche Médicale, and Ministero Istruzione, Università e Ricerca to interface contract with Paris V University (Y.B.-A.). We are grateful to Drs. K. Kaila, P. Legendre, N. Burnashev, G. Chazal, and I. Medina for suggestions and critical remarks. We are grateful to Drs. L. Aniksztejn, F. Libersat, I. Bureau, and J. Epszstein for supervision of the results obtained by the Institut de Neurobiologie de la Méditerranée teams.

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DOI:10.1523/JNEUROSCI.3314-10.2011

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We report that physiological plasma levels of D-BHB, lactate, and pyruvate are in pups (P5–P7) 0.9, 1.5, and 0.12 mM, respectively. Then, using a wide range of techniques that include extracellular field potential, cell-attached single-channel, and perforated patch-clamp recordings and calcium imaging we report that neither DL-BHB (Sigma-Aldrich) nor physiological concentrations of pyruvate alter GABA actions or spontaneous network dynamics, notably GDPs on rat neocortical and hippocampal neurons. Only very high nonphysiological concentrations of pyruvate altered GABA signaling and GDPs. Our results suggest that depolarizing GABA and GDPs are attributable to neither the absence of BHB/pyruvate nor the metabolic state of neurons in glucose-containing ACSF.

Materials and Methods

All investigations were analyzed in a double-blind manner with results obtained by an investigator analyzed by another researcher. In addition, this study was supervised by an independent group of Institut de Neurobiologie de la Méditerranée (INMED) principle investigators that were not involved in research on GABA in the developing brain (see Acknowledgments). They critically reviewed the results that were discussed in internal meetings of all INMED researchers.

Endogenous plasma D-BHB, lactate, and pyruvate

For D-BHB determination, plasma of pups was first deproteinized using 6% (w/v) perchloric acid and centrifuged, and the supernatant was neutralized with KOH before enzymatic determination. D-BHB was determined enzymatically using the spectrophotometric procedure as described previously (Ferré et al., 1983). Dosage of serum lactate and pyruvate were done in the metabolic biochemistry laboratory of Timone Hospital (Marseille, France). Enzymatic technique was used for lactate (RAPIDLAB 1265). Enzymatic dosage based on the reduction of pyruvate to lactate by the lactate dehydrogenase at pH 7.5 with excess nicotinamide adenine dinucleotide was used for the dosage of pyruvate (Vassault, 1991).

Brain slices

Brain slices were prepared from P4–P8 Wistar rats of both sexes. All animal use protocols conformed to the national guidelines on the use of laboratory animals and were approved by the Animal Care and Use Committees of Inserm and International School for Advanced Studies. Animals were rapidly decapitated, and brains were removed. Coronal slices (300–500 μ m) were cut using a tissue slicer (Leica-VT1200S; Microm International) in ice-cold oxygenated modified ACSF with 0.5 mM CaCl_2 and 7 mM MgSO_4 , in which Na^+ was replaced by an equimolar concentration of choline. Slices were then transferred to oxygenated (95% O_2 /5% CO_2) standard ACSF containing the following (in mM): 126 NaCl, 3.5 KCl, 2.0 CaCl_2 , 1.3 MgCl_2 , 25 NaHCO_3 , 1.2 NaH_2PO_4 , and 10 glucose, pH 7.4, at room temperature (20–22°C) for at least 1 h before use. For recordings, slices were placed into a conventional, fully submerged chamber superfused with ACSF (32–34°C) at a rate of 2–3 ml/min.

Perforated patch-clamp and whole-cell recordings

Patch-clamp recordings were performed from neocortical pyramidal and CA3 pyramidal neurons using EPC-10 Double (HEKA Elektronik Dr. Schulze GmbH) and Axopatch 200A (Molecular Devices) amplifiers. Patch electrodes were made from borosilicate glass capillaries (GC150F-15; Clark Electromedical Instruments). Patch pipette solution for gramicidin perforated patch-clamp recording contained the following (in mM): 150 KCl and 10 HEPES, buffered to pH 7.2 with Tris-OH. Gramicidin was first dissolved in DMSO to prepare a stock solution of 10–40 mg/ml and then diluted in the pipette solution to a final concentration of 80 μ g/ml. The gramicidin-containing solution was prepared and sonicated <1 h before the experiment. To facilitate cell-attached formation (4–10 G Ω), patch pipettes were backfilled with a gramicidin-containing solution. Between 20 and 30 min after formation of the cell-attached seal, the series resistance (R_s) stabilized at 8–60 M Ω . Series resistance was monitored during all recording sessions. At the end of each recording,

negative pressure was applied to break the membrane and establish whole-cell configuration. This was associated with a shift of the reversal potential of the GABA-mediated responses to near 0 mV. The membrane potential values (E_m) were corrected for series resistance offline as $V(\text{corrected}) = V(\text{holding}) - IR_s$. For whole-cell recordings, we used the pipette solution containing the following (in mM): 135 K-gluconate, 20 KCl, 10 HEPES, 4 MgATP, 0.3 GTP, and 0.5 EGTA. A picospritzer (General Valve Corporation) was used to puff apply GABA (100 μ M in ACSF) from a glass pipette in stratum radiatum at a distance of $\sim$ 100 μ m from the soma in gramicidin perforated patch recordings. The pressure varied from 10 to 20 kPa, and the duration of the puff varied from 50 to 200 ms.

Cell-attached recordings of GABA_A and NMDA receptor channels

Patch-clamp recordings from visually identified pyramidal cells in a cell-attached configuration were performed using an EPC-10 double amplifier or Axopatch 200B amplifier. For recordings of single GABA channels, the following patch pipette solution was added on the day of the experiment from a 1 mM frozen stock solution (in mM): 120 NaCl, 5 KCl, 20 tetraethylammonium-Cl, 5 4-aminopyridine, 0.1 CaCl_2 , 10 MgCl_2 , 10 glucose, and 10 HEPES-NaOH, buffered to pH 7.2–7.3 (with GABA at 1–5 μ M). E_m was estimated using cell-attached recordings of single NMDA receptor (NMDAR) channels as described previously (Tyzio et al., 2003). For recordings of single NMDAR channels, pipette solution contained nominally Mg^{2+} -free ACSF with NMDA (10 μ M), glycine (1 μ M), and strychnine (1 μ M). Pipettes (resistance of 3.5–8 M Ω) were pulled from borosilicate glass capillaries (GC150F-15; Clark Electromedical Instruments). Recordings were digitized (10 kHz) online with Digidata 1200 or 1440 interface cards (Molecular Devices), filtered (2.9 kHz), and analyzed offline with Axon package (Molecular Devices) and Origin (Microcal Software) as described previously (Tyzio et al., 2003, 2006). Group measures are expressed as means $\pm$ SEM; error bars also indicate SEM. The statistical significance of differences was assessed with Student's *t* test. The level of significance was set at $p < 0.05$.

Calcium imaging

Slice preparation for calcium imaging. Horizontal slices of neocortex and hippocampus (400 μ m thick) were prepared from P7 rats using a Vibratome tissue slicer (Leica VT 1200S) in ice-cold oxygenated modified ACSF (mACSF) (with 0.5 mM CaCl_2 and 7 mM MgSO_4 ; NaCl replaced by an equimolar concentration of choline). Slices were then transferred for rest ($\sim$ 1 h) in oxygenated normal ACSF containing the following (in mM): 126 NaCl, 3.5 KCl, 1.2 NaH_2PO_4 , 26 NaHCO_3 , 1.3 MgCl_2 , 2.0 CaCl_2 , and 10 D-glucose, pH 7.4. For AM loading, slices were incubated in a small vial containing 2.5 ml of oxygenated ACSF with 25 μ l of a 1 mM fura-2 AM solution (in 100% DMSO; Invitrogen) for 20–30 min. Slices were incubated in the dark, and the incubation solution was maintained at 35–37°C. Slices were perfused at a rate of 4 ml/min with continuously aerated (95% O_2 /5% CO_2) normal ACSF at 35–37°C. Imaging was performed with a multibeam two-photon laser scanning system (Triscope-LaVision Biotec) coupled to an Olympus microscope as described previously (Crépel et al., 2007). Images were acquired through a CCD camera (La Vision Imager 3QE), which typically resulted in a time resolution of $\sim$ 100 ms (2 $\times$ 2 binning; pixel size, 600 nm). Slices were imaged using a low-magnification, high numerical aperture objective (20 $\times$, numerical aperture 0.95; Olympus). The size of the imaged field was typically 430 $\times$ 380 μ m². Imaging depth was on average 80 μ m below the surface (range, 50–100 μ m).

Analysis of multineuron calcium activity. Analysis of the calcium activity was performed using a previously designed software for neocortical and hippocampal slice analysis (Allène et al., 2008). To summarize briefly, this allowed (1) automatic identification of loaded cells, (2) measuring the average fluorescence transients from each cell as a function of time, and (3) detecting the onsets and offsets of calcium signals. To quantify synchronous activity patterns, we used two parameters: frequency and amplitude and duration of synchronous events. The frequency of a network pattern was derived from the average time interval between two peaks of synchronous activity. The amplitude of a network pattern in a given movie was the average of the maximum of cells coactive in each peak of synchrony across the movie. To identify peaks of synchro-

nous activity that included more cells than expected by chance, we used interval reshuffling (randomly reordering of intervals between events for each cell) to create set of surrogate event sequences. Reshuffling was performed 1000 times for each movie, and a surrogate histogram was constructed for each reshuffling. The threshold corresponding to a significance level of $p < 0.05$ was estimated as the number of coactive cells exceeded in a single frame in only 5% of these histograms. This threshold was used to calculate the duration of a synchronous activity pattern that is the number of successive frames for which the number of coactive cells was superior to threshold. Experimental values are given as means $\pm$ SEMs. Student's t test and χ^2 test were used for statistical comparisons.

Chemical identification of contaminants

One milligram of DL-BHB was dissolved in a mixture of ethyl acetate and pentafluoropropionic anhydride (PFPA) (Aldrich) and composed of 80 μ l each. After 30 min at 80°C, 1 μ l of the mixture was directly injected into a gas chromatography–mass spectrometry (GC–MS) instrument (Agilent Technologies) equipped with a 6890N GC and a 5973 MSD system.

Electron impact mass spectrum of dibenzylamine (DBA) as PFPA derivative was identified in the DL-BHB acid standard from Acros Organics but not in DL-BHB from Sigma-Aldrich. A PFPA was identified as its molecular ion at a mass-to-charge ratio (m/z) of 383 and a typical fragmentation with the predicted M-91 at m/z of 252 and the benzyl core at m/z of 91. Chromatographic conditions were as follows: DB-1MS capillary column (Agilent Technologies), 30 m $\times$ 0.25 mm inner diameter, film thickness of 1.0 μ m; column temperature, 100–260°C at a rate of 4°C/min; injection port, 280°C; gas vector helium at 1 ml/min (10.4 psi); electron impact mode at an ionization energy of 70 eV. Comparisons of mass spectra were done by using a standard of DBA treated in the same conditions.

Pharmacology

Bicuculline, 2,3-dihydroxy-6-nitro-7-sulfonyl-benzo[*f*]quinoxaline, D-APV, DNQX, and CNQX were purchased from Tocris Bioscience; DL-BHB was from Sigma-Aldrich (catalog #54965, batch #1316259 31908044) and from Acros Organics. Isoguvacine, dybenzylamine, GABA, lactate, and pyruvate were from Sigma. TTX was from Ascent Scientific Ltd.

Results

Blood concentrations of D-BHB, lactate, and pyruvate in neonatal rodent

In newborn rodents, the blood levels of active D-BHB are in the submillimolar range (see Discussion). We reinvestigated this issue, using an assay that determines D-BHB levels, and found that, in P7 rats, the D-BHB plasma concentration was 0.91 ± 0.15 mM ($n = 14$). We measured D-BHB levels after administration of insulin (0.33 U/kg, i.m.) to inhibit fatty acid oxidation and 2-mercaptoacetate (100 mg/kg, i.p.) to inhibit mitochondrial acyl-CoA dehydrogenase with 5 mM glucose to prevent hypoglycemia. This treatment strongly and rapidly (within 1 h) reduced the plasma level of the D-BHB to 0.14 ± 0.03 mM ($n = 14$). Conversely, injections of exogenous 4 mM DL-BHB increased the blood D-BHB concentration to 1.74 ± 0.42 mM ($n = 7$). Using conventional hospitals kits, plasma lactate and pyruvate levels at P4 were of 1.5 ± 0.25 mM and 123 ± 16 μ M, respectively ($n = 5$). Therefore, plasma D-BHB, pyruvate, and lactate concentrations are much lower than those used by Zilberter and colleagues (4

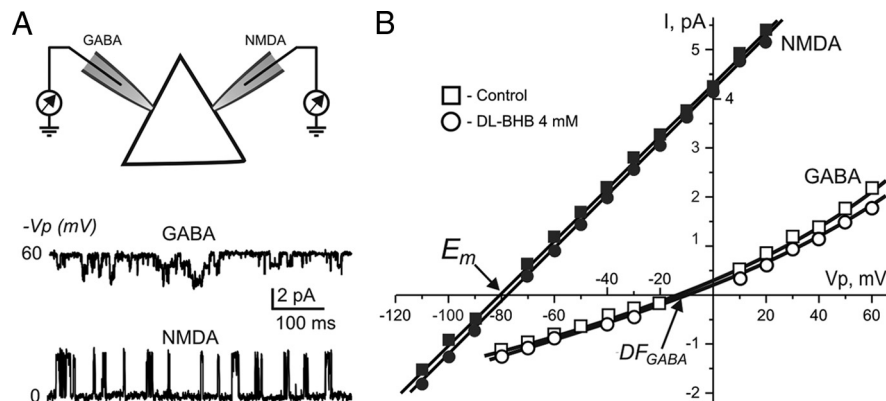


Figure 1. Estimation of E_{GABA} by double recordings of DF_{GABA} and E_m from the same neocortical cell. **A**, Scheme of consecutive cell-attached recordings of single NMDAR and GABA_AR channels. First we recorded from the same neuron in control conditions for NMDAR channels (for E_m) and then GABA_AR channels (for DF_{GABA}). These two measurements allow us to determine E_{GABA} for this neuron. Then, we applied DL-BHB for 40 min and repeated the recordings with the same sequence (NMDAR and then GABA_AR channels). Thus, every cell studied was patched four times. Representative traces of recordings of the single-channel openings are shown below. **B**, Representative plot of I – V relationships of single GABA_AR and NMDAR channels used for estimation of E_{GABA} in neocortical pyramidal cell ($E_{\text{GABA}} = DF_{\text{GABA}} + E_m$). Each point is mean amplitude of ~ 30 openings at a given pipette potential (V_p). The reversal potential that corresponds to DF_{GABA} was estimated by the exponential growth fit of the I – V curve. The current–voltage relationships of NMDAR channels were best fitted with linear function (Tyzio et al., 2003, 2008). Note that application of DL-BHB does not change significantly DF_{GABA} and E_m .

mM DL-BHB, 5 mM pyruvate, and 5 mM lactate). The difference is particularly high for pyruvate (40-fold) (see Discussion).

DL-BHB (from Sigma-Aldrich) does not alter the driving force for somatic GABA_AR-mediated currents

The binding of GABA to GABA_A receptors opens channels permeable notably to chloride. The resulting trans-membrane chloride current can either depolarize or hyperpolarize the membrane according to its E_{GABA} and E_m of the cell. As indicated in previous studies, measurement of E_m using NMDAR channels as the voltage sensor has an advantage over the other microelectrode techniques especially in immature cells with high input resistance (Tyzio et al., 2003). Single NMDAR and GABA_AR channels were recorded from either the same neurons or from different neurons to determine E_m and driving force for somatic GABA_AR-mediated currents (DF_{GABA}) and thereby allow a precise determination of E_{GABA} (Tyzio et al., 2008). As in the studies by Rheims et al. (2009) and Holmgren et al. (2010), slices were incubated in 4 mM DL-BHB (Sigma-Aldrich) for at least 40 min and then transferred to a recording chamber in which they were continuously superfused with the same concentration of DL-BHB.

We first determined E_{GABA} by alternate cell-attached recordings (in sum, four attempts for every cell) of NMDAR and GABA_AR single channels from the soma of the same neuron in the neocortex (Fig. 1). As shown in Table 1, DL-BHB altered neither E_m nor E_{GABA} in neocortical neurons (P4–P5 rats) of both superficial and deep layers (all $p > 0.05$). Similar results were obtained when single NMDAR and GABA_AR channels were recorded in different neurons (to avoid the potential local membrane alterations): DF_{GABA} in hippocampal CA3 pyramidal cells was not significantly different in control and 4 mM DL-BHB (Table 2) ($p > 0.05$); E_m in the same population of cells also did not change significantly in DL-BHB ($p > 0.05$). Similarly, in both superficial and deep layers of the neocortex, DF_{GABA} was strongly depolarizing in control conditions and in the presence of DL-BHB; the values of E_m were similar in control conditions and in the presence of DL-BHB (all $p > 0.05$) (Table 2). Therefore, DL-

Table 1. Double measurement of DF_{GABA} and E_m from the same neocortical cell reveal that DL-BHB does not alter E_{GABA} in pyramidal neocortical cells

	Regular ACSF	ACSF + DL-BHB at 4 mM
<i>n</i>	6	6
E_m (mV)	-81.7 ± 2.6	-79.2 ± 2.9
DF_{GABA} (mV)	14.8 ± 3.3	13.7 ± 3.1
E_{GABA} (mV)	-66.9 ± 3.9	-65.5 ± 3.1

DF_{GABA} inferred from $I-V$ curves of single $GABA_A$ R channels, E_m inferred from $I-V$ curves of single NMDAR, and the $GABA_A$ R channels reversal potential.

BHB alters neither E_m nor E_{GABA} in neocortical and hippocampal neurons.

DL-BHB does not alter the polarity of synaptic GABAergic responses

We then used perforated patch recordings to determine the effects of DL-BHB on synapse-driven $GABA_A$ R-mediated postsynaptic potentials (GPSPs). In immature hippocampal slices, GABA released from MF terminals exerts a depolarizing action on CA3 principal cells in gramicidin perforated patch recordings (Sivakumaran et al., 2009). With gramicidin patch recordings from CA3 pyramidal cells, the resting membrane potential was -56 ± 2.2 mV ($n = 10$) (Fig. 2*A–C*). The more depolarized E_m value found with these experiments (with respect to that observed in cell-attached recordings) could be attributed to the leakage via gigaseal contact introduced in perforated patch recordings (Barry and Lynch, 1991; Tyzio et al., 2003). In the presence of 20 μ M DNQX and 50 μ M D-AP-5 to block AMPA and NMDA receptors, respectively, local stimulation of GABAergic interneurons in stratum radiatum generated PSPs with reversed polarity (E_{GPSPs}) at -47.6 ± 3.3 mV ($n = 10$). The driving force for GABA was ~ 9 mV positive to E_m , indicating that GABA is depolarizing from the resting potential as a result of the activity of chloride accumulating NKCC1 cotransporter (Tyzio et al., 2003, 2007; Sipila et al., 2006). Addition of DL-BHB (4 mM for at least 40 min) altered neither E_m (-60 ± 1 mV) nor E_{GPSPs} (-48 ± 2 mV; $n = 11$), with an average DF_{GABA} of ~ 12 mV. The values of E_m and E_{GPSPs} obtained in the presence of DL-BHB were not significantly different from those obtained in the absence of DL-BHB ($p > 0.05$ for both) (Fig. 2*A–C*). Therefore, DL-BHB does not alter E_{GPSPs} .

We also used gramicidin perforated patch recordings to determine whether DL-BHB alters the response evoked by $GABA_A$ receptor agonists. The reversal of isoguvacine-evoked responses (E_{iso}) was determined in control and during bath application of 4 mM DL-BHB for at least 40 min. Isoguvacine (40 μ M, from a holding potential of -60 mV) generated inward currents that reversed at -48 ± 3 mV in control ($n = 6$) and -47 ± 3 mV in the presence of DL-BHB ($n = 6$). These values were not significantly different ($p > 0.05$) (Fig. 2*D*).

Furthermore, in three cells kept in current-clamp conditions in the presence of blockers of fast synaptic transmission (DNQX at 50 μ M, D-APV at 50 μ M, and bicuculline at 10 μ M), bath application of 4 mM DL-BHB altered neither the resting membrane potential nor the input resistance [values for E_m and R_{in} were -60 ± 3 and -61 ± 2 mV ($n = 6$) and 625 ± 37 and 575 ± 42 M Ω ($n = 6$, all $p > 0.05$), before and 40 min after DL-BHB application, respectively] (supplemental Fig. 1, available at www.jneurosci.org as supplemental material).

DL-BHB does not alter the excitatory effects of GABA

To determine whether DL-BHB alters the excitatory actions of GABA, we focally applied GABA on neurons recorded in cell-

attached configuration in the presence of CNQX (10 μ M), APV (40 μ M), and CGP 55845 ((2*S*)-3-[[[(1*S*)-1-(3,4-dichlorophenyl)ethyl]amino-2-hydroxypropyl](phenylmethyl)phosphinic acid hydrochloride) (2 μ M) to block ionotropic glutamate and metabotropic $GABA_B$ receptors.

In these conditions, GABA triggered spikes and this action was reversibly blocked by the $GABA_A$ receptor antagonists (data not shown), indicating that they were generated by the activation of $GABA_A$ receptors. As shown in Figure 3*A*, focal applications of GABA, in the hippocampus in the presence of DL-BHB generated a similar number of spikes as in controls (1.29 ± 0.09 spikes in control and 1.44 ± 0.1 in DL-BHB, $n = 7$, $p > 0.05$). Similar results were obtained with neocortical layer V pyramidal neurons (1.39 ± 0.06 spikes in control and 1.23 ± 0.05 in the presence of DL-BHB, $n = 6$, $p > 0.05$). The specific NKCC1 antagonist bumetanide (10 μ M) prevented GABA from inducing spikes generated by focal applications of GABA in the presence of the same mixture and DL-BHB, confirming that they were generated by depolarizing actions of GABA (Fig. 3*B*) ($n = 3$). Therefore, DL-BHB applied in conditions similar to those used by Rheims et al. (2009) and Holmgren et al. (2010) does not alter the driving force and reversal potential of somatic (extrasynaptic) and synaptic GABA activated responses, the resting membrane potential, and excitatory actions of GABA on neocortical or hippocampal neurons.

DL-BHB does not alter GDPs

We next examined whether DL-BHB affects GDPs, which are dependent on depolarizing actions of GABA and are very sensitive to alterations of neuronal excitability and to insufficient energy supply (Ben-Ari et al., 1989; Dzhalal et al., 1999; Allène et al., 2008). As shown in Figure 4, GDPs occurred synchronously with extracellular field potentials and were readily identified in whole-cell recordings by their characteristic shape and kinetics. DL-BHB altered neither the frequency [0.054 ± 0.005 and 0.046 ± 0.004 Hz in control ($n = 7$) and DL-BHB ($n = 8$), respectively; $p > 0.05$] nor the integrated area under GDPs [26.6 ± 3.1 mV/s ($n = 6$) and 24.3 ± 3.2 mV/s ($n = 7$) in control and DL-BHB, respectively; $p > 0.05$].

We next used fast multineuron calcium imaging to measure the actions of DL-BHB on neuronal activity synchronization in cortical networks. This approach enables investigating the spatio-temporal dynamics of spontaneous neuronal activity in large neocortical and hippocampal networks with single-cell resolution. With this approach, immature hippocampal and neocortical networks display similar correlated activity patterns composed of synchronous plateau assemblies (SPAs) and GDPs that are both strongly modulated by the excitatory action of GABA (Crépel et al., 2007; Allène et al., 2008; Bonifazi et al., 2009). These patterns can be easily identified in single cells based on their characteristic calcium dynamics because SPAs are associated with long-lasting (~ 10 – 20 s on average) calcium plateau potentials synchronized across small groups of neurons, whereas the intracellular calcium correlate of GDPs are fast calcium transients (~ 250 ms) decay (Crépel et al., 2007; Allène et al., 2008) synchronizing larger neuronal populations. To determine whether DL-BHB affected neuronal calcium dynamics, multibeam two-photon imaging in neocortical slices loaded with a calcium indicator (fura-2 AM) was performed. Slices were incubated in 4 mM DL-BHB for at least 40 min. Custom software was used to measure fluorescence changes in each cell and to mark the onset and offset of individual calcium transients (see Materials

Table 2. DL-BHB does not alter DF_{GABA} and E_m in principal cells of rat hippocampus and neocortex: changes of DF_{GABA} determined with cell-attached recording of $GABA_A$ R channels in hippocampal CA3 cells and both superficial and deep layers of neocortex

	Hippocampus		Neocortex	
	regular ACSF	ACSF + DL-BHB at 4 mM	regular ACSF	ACSF + DL-BHB at 4 mM
E_m (mV)	-77.8 ± 3.5 ($n = 9$)	-76.2 ± 4.2 ($n = 12$)	-72.2 ± 5.4 ($n = 5$)	-70.6 ± 3.2 ($n = 7$)
DF_{GABA} (mV)	19.8 ± 6.5 ($n = 12$)	16.5 ± 3.5 ($n = 14$)	22.9 ± 4.2 ($n = 10$)	18.2 ± 3.6 ($n = 11$)

Note that, in this set of experiments, DF_{GABA} and E_m were examined in different cells.

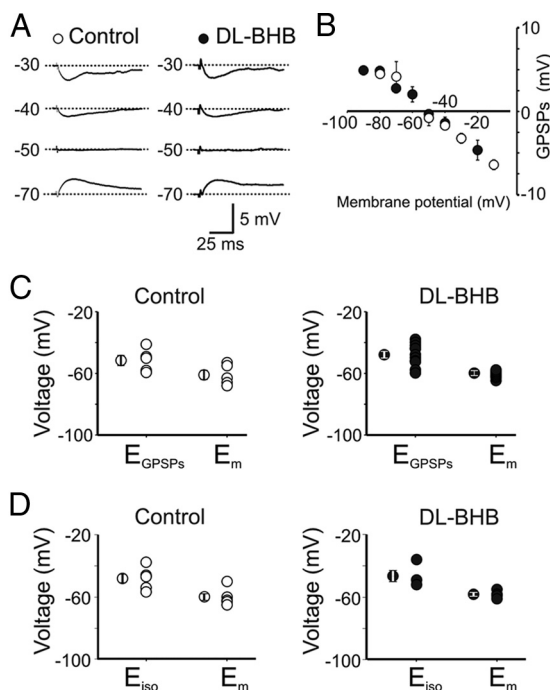


Figure 2. DL-BHB does not alter the polarity of GABAergic responses in CA3 pyramidal cells. **A**, Example of $GABA_A$ -mediated postsynaptic potentials evoked in control and in the presence of 4 mM DL-BHB at four different holding potentials (to the left of the traces) by local stimulation of $GABA_A$ ergic interneurons in stratum radiatum. **B**, The mean GPSP amplitudes obtained in 11 cells are plotted against membrane potentials (E_m). Vertical bars represent the SEM. **C**, Each symbol represents the E_{GPSPs} and the E_m of individual cells. Average values are shown on the left of each group (control: $E_{GPSPs} = -47.6 \pm 3.3$ mV, $E_m = -56 \pm 2.2$ mV, $n = 10$; in DL-BHB: $E_{GPSPs} = -48.2 \pm 2$ mV, $E_m = -60 \pm 1$ mV, $n = 11$). **D**, The E_{iso} was determined in control and during bath application of DL-BHB (4 mM, for at least 40 min). The isoguvacine application (40 μ M) generated the responses, which reversed at -48 ± 3 mV in control ($n = 6$) and -47 ± 3 mV in DL-BHB ($n = 6$), respectively. These values were not significantly different ($p > 0.05$).

and Methods). In these conditions, we did not find any significant decrease in either the fraction of neurons producing SPAs or the frequency and amplitude of GDPs in neocortical slices ($p > 0.05$, $n = 5$ slices) (Fig. 5, Table 3). As a positive control for excitatory GABA actions, we applied the NKCC1 antagonist bumetanide (10 μ M). Bumetanide significantly reduced the occurrence of GDPs both in regular ACSF and in the presence of DL-BHB (Fig. 5). Frequency and amplitude of GDPs were reduced to 26 ± 21 and $38 \pm 10\%$, respectively, in regular ACSF ($n = 7$, $p < 0.05$) and to 12 ± 7 and $56 \pm 34\%$, respectively, in DL-BHB conditions ($n = 5$, $p < 0.05$). As reported previously in hippocampal slices (Crépel et al., 2007), the fraction of SPA cells was significantly increased in the presence of bumetanide (10 μ M, to 535 ± 192 and $349 \pm 104\%$ in regular ACSF and DL-BHB, respectively, $n = 5$, $p < 0.05$). The effects of bumetanide cannot be reconciled with a reduction of the depolarizing action of GABA in the presence of DL-BHB (Rheims et al., 2009) (see Discussion).

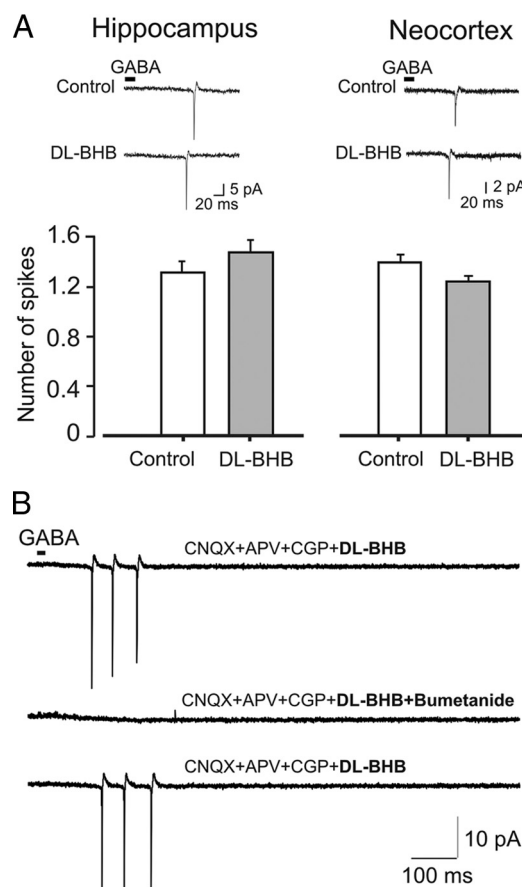


Figure 3. DL-BHB does not alter the excitation produced by focal application of GABA in both hippocampus and neocortical pyramidal neurons. **A**, Each column represents number of spikes induced by focal application of GABA (top traces) recorded in cell-attached mode from hippocampus and neocortex in control and in presence of 4 mM DL-BHB. **B**, Bumetanide (10 μ M) fully blocked the spikes generated by focal applications of GABA in the presence of the same mixture and DL-BHB ($n = 3$).

A contamination in DL-BHB (from Acros Organics) reduces DF_{GABA}

While conducting our experiments, we found that DL-BHB from another source (Acros Organics) gave different results. In slices incubated in 4 mM DL-BHB (Acros Organics for at least 40 min), DF_{GABA} measured using cell-attached recordings of $GABA_A$ R channels was significantly reduced in CA3 pyramidal cells from 12.2 ± 3.9 mV ($n = 6$) to 0.47 ± 3.04 mV ($n = 10$, $p < 0.05$; data not shown). Similar effects of DL-BHB (Acros Organics) were found with perforated patch recordings of GPSPs evoked by local stimulation of $GABA_A$ ergic interneurons in stratum radiatum in the presence of DNQX (20 μ M) and D-APV (50 μ M). As shown in supplemental Figure 2 (available at www.jneurosci.org as supplemental material), DL-BHB from Acros Organics (applied for at least 45 min) hyperpolarized the membrane and shifted E_{GPSPs} toward more negative values (on average, on seven neurons E_m

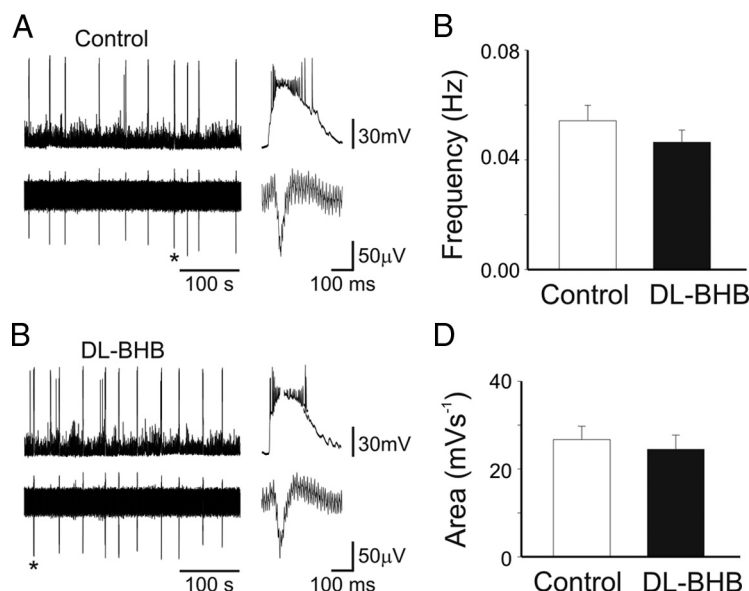


Figure 4. DL-BHB does not alter spontaneous neuronal activity patterns at early postnatal stages in hippocampal slices. **A, B**, Individual traces of spontaneous GDPs recorded at -70 mV from CA3 pyramidal cell (top traces) and field potentials (bottom traces) in slice exposed to ACSF (control, **A**) or ACSF plus DL-BHB at 4 mM. **B**, DL-BHB did not alter the frequency or the shape of GDPs (shown on the right in an expanded timescale). **C, D**, Each column represent the mean GDPs frequency (**C**) or area (**D**) in control (white; $n = 6$) or during bath application of DL-BHB (black; $n = 7$). $*p < 0.01$.

and E_{GPSPs} were -80 ± 4 and -67 ± 4 mV, respectively). In additional experiments, we tested the actions of DL-BHB (Acros Organics) on GDPs using calcium imaging. As shown in supplemental Figure 3 (available at www.jneurosci.org as supplemental material), DL-BHB (Acros Organics) reduced the frequency and amplitude of GDPs in neocortical slices (from 0.12 ± 0.02 to 0.02 ± 0.01 Hz and from 22 ± 3 to $5 \pm 4\%$ of active cells, $n = 5$, $p < 0.05$). Therefore, DL-BHB (Acros Organics) reduces DF_{GABA} and alters GDPs.

Because previous studies reported the presence of a contaminant in L-BHB (Donevan et al., 2003) (see Discussion), we decided to test whether a similar contaminant was also present in DL-BHB obtained from this source (Acros Organics). Using a GC-MS instrument, several contaminants were found in this source of DL-BHB, notably DBA as PFPA derivative identified in the DL-BHB from Acros Organics (supplemental Fig. 4, available at www.jneurosci.org as supplemental material) but not in DL-BHB (Sigma-Aldrich; data not shown). We therefore tested the actions of DBA on DF_{GABA} and found that $50 \mu\text{M}$ DBA switched DF_{GABA} , determined with single-channel recordings, from depolarizing 12.8 ± 2.8 mV ($n = 10$) to hyperpolarizing -2.9 ± 2.2 mV ($n = 10$, $p < 0.001$; data not shown). With whole-cell recordings, DBA also blocked GDPs in a concentration-dependent way with an EC_{50} of $57 \mu\text{M}$ (supplemental Fig. 5, available at www.jneurosci.org as supplemental material). The dose-response curve was steep: whereas at $60 \mu\text{M}$ DBA severely reduced the frequency of GDPs from 0.046 ± 0.007 to 0.01 ± 0.002 Hz, at $80 \mu\text{M}$ it completely abolished them ($n = 10$). These observations suggest that DL-BHB does not alter GABA signals, but care must be taken when using BHB compounds to ensure absence of contaminants. We next investigated the actions of pyruvate on the same parameters.

High but not physiological concentrations of pyruvate affect GABA signaling

We first tested the effect of pyruvate at 5 mM on DF_{GABA} in single-channel recordings of CA3 pyramidal cells from P7 rat hip-

pocampus. DF_{GABA} shifted from 12.2 ± 5.7 mV in control ($n = 9$) to 0.4 ± 3 mV in pyruvate ($n = 9$, $p < 0.01$; data not shown). In contrast, more relevant physiological concentrations ($200 \mu\text{M}$) did not significantly change DF_{GABA} (7.1 ± 3.2 mV in control, $n = 7$ and 6.7 ± 2.5 mV in pyruvate, $n = 8$, $p > 0.05$; data not shown). Therefore, pyruvate does not affect DF_{GABA} at physiological levels and alters DF_{GABA} only at excessively high concentrations.

Gramicidin perforated patch experiments were performed to assess whether pyruvate alters E_m and E_{GPSPs} . Like DL-BHB, the addition of a physiological concentration of $200 \mu\text{M}$ pyruvate to the ACSF did not modify E_m and E_{GPSPs} (Fig. 6). E_m values were -56.8 ± 0.9 and -57.1 ± 1.02 mV in control and in the presence of pyruvate, respectively ($p > 0.05$, $n = 7$). In contrast, 5 mM pyruvate caused a negative shift of E_{GPSPs} (from -48.3 ± 1.9 to -55.7 ± 3 mV, $n = 7$) without altering E_m (E_m values were -56.8 ± 0.9 and -56.6 ± 1.8 mV in control and in the presence of 5 mM pyruvate, respectively, $n = 7$). The driving force for GABA was ~ 0.9 mV positive to E_m , indicating that GABA does not exert a depolarizing action. The E_{GPSPs} value observed in the presence of 5 mM pyruvate was significantly different from that obtained in control ($p = 0.003$) or in the presence of $200 \mu\text{M}$ pyruvate ($p = 0.005$). E_m and E_{GPSPs} were unaffected by the further addition of DL-BHB to 5 mM pyruvate. In the presence of 4 mM DL-BHB and 5 mM pyruvate, E_m and E_{GPSPs} were -57.4 ± 2.5 and -59.6 ± 2.9 mV, respectively ($n = 6$). These values are similar to those obtained when cells were exposed only to pyruvate (data not shown).

Furthermore, with extracellular field potential recordings from the CA3 region (P4–P7), pyruvate at a physiological concentration did not affect the frequency of GDPs (0.06 ± 0.008 and 0.06 ± 0.01 Hz in control and in the presence of $200 \mu\text{M}$ pyruvate, respectively; $p > 0.05$, $n = 6$) (Fig. 7). In contrast, in the presence of 5 mM pyruvate, the frequency of GDPs severely depressed (from 0.06 ± 0.008 to 0.02 ± 0.007 Hz, $p = 0.005$, $n = 6$).

Similar observations were made with imaging techniques. We monitored spontaneous neuronal activity in neocortical and hippocampal slices bathed in mACSF that mimics physiological conditions (lactate at 1.5 mM, pyruvate at $150 \mu\text{M}$, and DL-BHB at 2 mM; see Discussion). There was no change in GDPs or SPA patterns of activity (Fig. 8, Table 4) ($p > 0.05$, five neocortical slices and three hippocampal slices). Indeed, the frequency of neocortical GDPs in mACSF was 0.1 ± 0.02 Hz compared with 0.12 ± 0.02 Hz in control conditions, and the amplitude of neocortical GDPs was 24 ± 4 compared with $22 \pm 3\%$ of active cells in regular ACSF. The frequency of hippocampal GDPs in mACSF was not affected either (0.07 ± 0.03 vs 0.1 ± 0.03 Hz in control conditions). In contrast, pyruvate (5 mM), added to regular ACSF, significantly reduced the frequency and amplitude of hippocampal GDPs to 8 ± 7 and $53 \pm 33\%$ of control conditions, respectively (supplemental Fig. 6) ($n = 5$, $p < 0.05$). Therefore, pyruvate alters GABA postsynaptic currents and GDPs only at high nonphysiological concentrations.

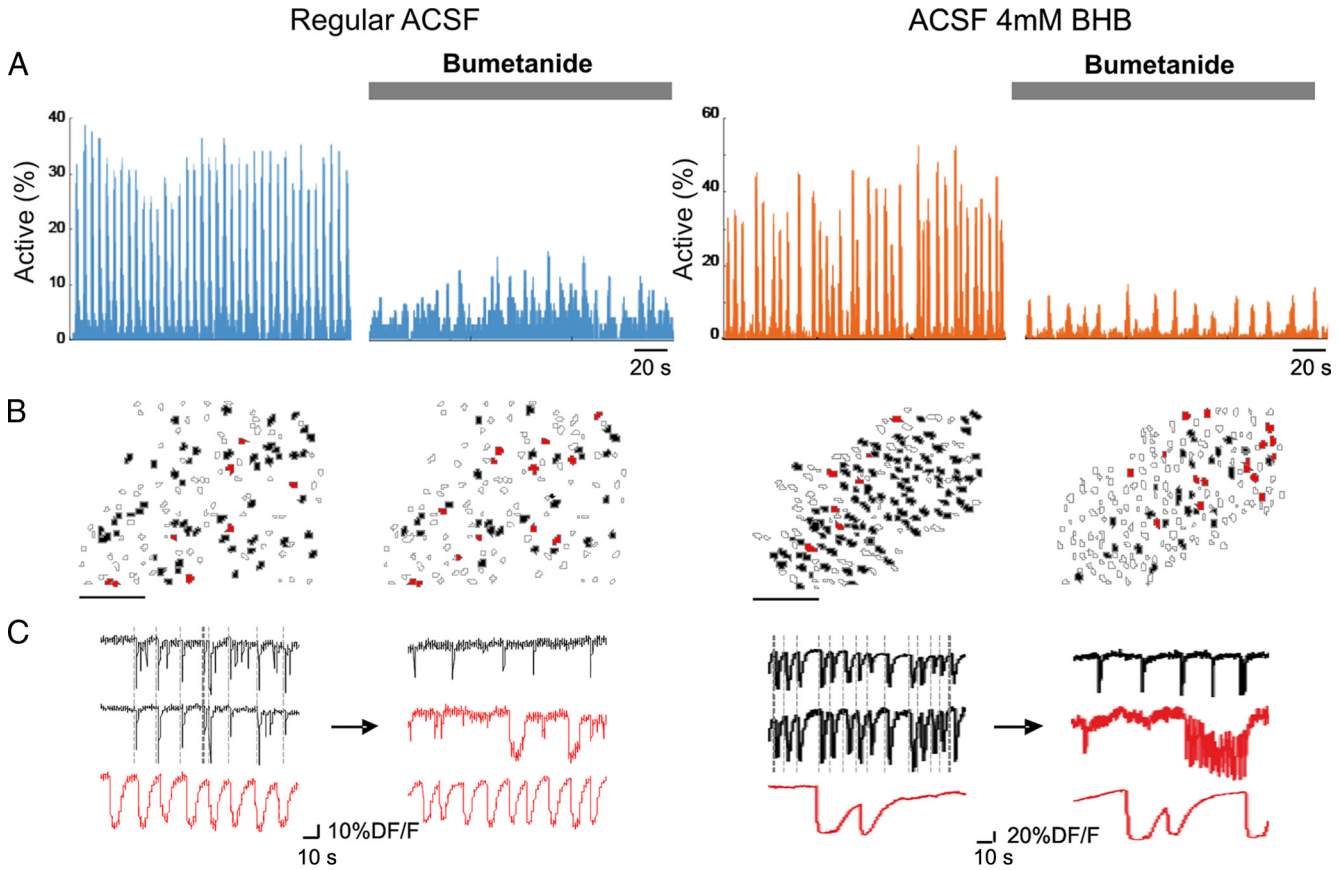


Figure 5. DL-BHB does not alter spontaneous neuronal activity patterns in neocortical slices. **A**, Histograms indicating the fraction of active cells as a function of time in calcium movies in regular ACSF and in the presence of 4 mM DL-BHB (Sigma-Aldrich). Each peak of synchronous neuronal activity in the histograms corresponds to a GDP. GDPs were strongly reduced in the presence of the NKCC1 antagonist bumetanide (10 μ M) in neocortical slices from P7 rats. **B**, Automatically detected contours of the imaged cells: open contours indicate silent cells, black filled contours indicate cells involved in GDPs, and red filled contours are SPA cells. Note that the number of SPA cells relative to the number of active cells increased in the presence of 10 μ M bumetanide in the neocortex and hippocampus (scale bar, 100 μ m). **C**, Calcium fluorescence traces of representative cells implicated in GDPs (black) and SPAs (red). Note that some GDP cells display an SPA pattern of activity after adding bumetanide (middle traces).

Table 3. Dynamics of cortical GDPs and SPAs in regular ACSF and in the presence of DL-BHB (Sigma-Aldrich)

	Regular ACSF	ACSF + DL-BHB at 4 mM
<i>n</i>	7	5
GDP _{freq} (Hz)	0.12 $\pm$ 0.02	0.14 $\pm$ 0.05
GDP _{amp} (%)	22 $\pm$ 3	22 $\pm$ 4
SPA cells (%)	8 $\pm$ 2	17 $\pm$ 11

GDP_{freq}, Frequency of occurrence of GDPs (in hertz); GDP_{amp}, fraction of active cells involved in GDPs (percentage); SPA cells, fraction of active cells involved in SPAs (percentage).

Discussion

We show that neither DL-BHB, used in similar concentrations as Zilberter and colleagues, nor pyruvate at physiologically relevant concentrations alters GABA depolarizing actions and GDPs, suggesting that depolarizing GABA and GDPs in immature neurons are not attributable “to energy deprived conditions when relying only on glucose” (Rheims et al., 2009; Holmgren et al., 2010).

Possible reasons for the discrepancies

DL-BHB at 4 mM (Sigma-Aldrich) had no effects on (1) E_m , E_{GABA} , and DF_{GABA} in somatic recordings of single GABA and NMDA channels, (2) E_m , synaptic GABAergic potentials and the responses evoked by GABA application in gramicidin perforated patch recordings, (3) spikes generated by GABA

in cell-attached recordings, and (4) frequency of GDPs or SPAs relying on calcium imaging and electrophysiological recordings.

The wide range of E_{GABA} values reported by Holmgren et al. (2010) suggests a heterogeneity possibly attributable to pooling different ages particularly between P1 and P4 in which major shifts occur in E_{GABA} (Tyzio et al., 2006, 2007) in superficial and deep neocortical layers neurons or CA1 and CA3 pyramidal neurons with their different age (Ben-Ari et al., 2007; Rheims et al., 2008). Additional explanations to the discrepancy between the present study and the work of Zilberter and colleagues include the experimental approaches to measure E_{GABA} . (1) Cell-attached recordings of GABA channels without potassium channel blockers in the pipette could be contaminated by potassium channels and shift the measures to potassium reversal. (2) Prolonged activation of GABA receptors by long (2 s) isoguvacine applications and voltage ramps at the peak of the response used to determine E_{GABA} can be associated with profound changes in ionic distribution. Here, we used brief (50–200 ms) isoguvacine applications and synaptic stimulations at different holding potentials to minimize the error. (3) The “random” burst protocol used to record the spikes evoked by synaptic activation of GABA receptors may introduce activity-dependent disturbance in ionic gradients (the same applies for repetitive puff application of isoguvacine). This was controlled in the present study by wash in

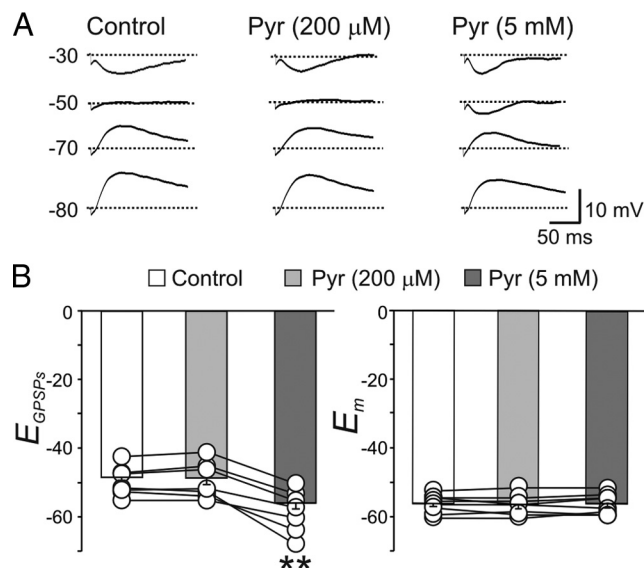


Figure 6. E_{GSPs} shift in the presence of high but not low concentrations of pyruvate. **A**, Examples of GABA_A-mediated postsynaptic potentials evoked in control (white, left), in the presence of 200 μ M pyruvate (Pyr) (light gray, middle), and in the presence of 5 mM pyruvate (dark gray, right). **B**, Open circles represent E_{GSPs} (graph on the left) and E_m (graph on the right) obtained in individual cells recorded in normal ACSF (white columns; left), in ACSF containing 200 μ M (light gray; middle) or 5 mM (dark gray; right) pyruvate ($n = 7$). Bars at the top of the columns represent the SEM. Note the negative shift of E_{GSPs} , but not in E_m , of cells exposed to 5 mM pyruvate. The mean E_{GSPs} value obtained in 5 mM pyruvate was significantly different from that obtained in control ($p = 0.003$) and in 200 μ M pyruvate ($p = 0.005$). ** $p < 0.01$.

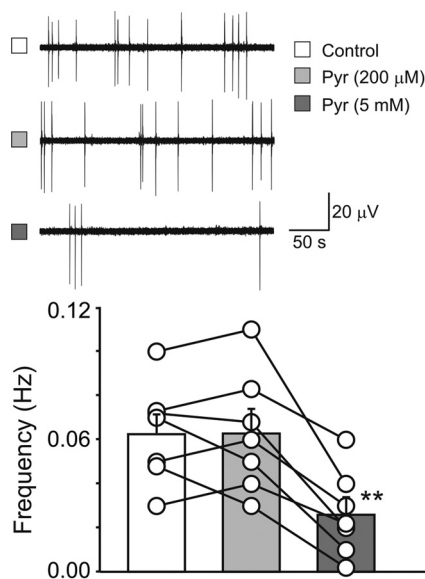


Figure 7. High but not low concentrations of pyruvate reduce GDPs frequency. On the left, sample traces of spontaneous GDPs recorded from the same slice before (white symbol; top trace) or during exposure to 200 μ M pyruvate (Pyr) (light gray; middle trace) and 5 mM pyruvate (dark gray; bottom trace). Note the reduction in GDPs frequency with 5 mM but not 200 μ M pyruvate. On the right, each symbol represents the mean frequency value of GDPs obtained in individual slices before (white column) or during exposure to 200 μ M (light gray) and 5 mM (dark gray; right) pyruvate ($n = 7$). Bars at the top of the columns represent the SEM. The mean GDPs frequency value obtained in 5 mM pyruvate was significantly different from that obtained in control ($p = 0.005$) and in 200 μ M pyruvate ($p = 0.005$). * $p < 0.05$.

and washout of bicuculline or bumetanide. Finally, the non-invasive quantitative determination of the percentage of neurons active during SPAs and GDPs suggests that both DL-BHB and pyruvate have no effects on these patterns.

The presence of a contaminant dibenzylamine in DL-BHB (Acros Organics) that changes DF_{GABA} and E_{GABA} is important for future studies. DBA mediates effects thought previously to be attributable to DL-BHB blockade of cardiac K^+ channels and anticonvulsive actions of ketone bodies (Doepner et al., 1997, 2001; Rho et al., 2002; Donevan et al., 2003). Investigations using DL-BHB must take this parameter into account.

Physiologically relevant concentrations of D-BHB, lactate, and pyruvate

Neonatal plasma concentrations of D-BHB (present results; see also Nehlig and Pereira de Vasconcelos, 1993; Lust et al., 2003; Vannucci and Simpson, 2003; Erecinska et al., 2004; Nehlig, 2004) are significantly lower than those used by Rheims et al. (2009) and Holmgren et al. (2010), particularly because only a fraction (40%) of plasma D-BHB is found in neonatal cortex (Lust et al., 2003). In addition, the argument that 4 mM DL-BHB is equivalent to 2 mM active D-BHB (Rheims et al., 2009) is invalidated by the observations that L-BHB exerts complex biological actions (Moore et al., 1976; Webber and Edmond, 1977; Herzberg and Gad, 1984; Eaton et al., 2003; Tsai et al., 2006; Chou et al., 2008). Also, ketogenic diet reduces glucose utilization by 10% per millimolar plasma ketone bodies (Robinson and Williamson, 1980; Harding and Charlton, 1990; LaManna et al., 2009), hampering the interpretation and relevance of observations made using ACSF with glucose and DL-BHB.

The lactate (5 mM) and pyruvate (5 mM) concentrations used by Holmgren et al. (2010) are never observed in postnatal physiological conditions. Plasma lactate levels are high *in utero* (10 mM), shift to 3 mM during the presuckling period, and remain thereafter close to 1 mM (Medina, 1985), and even then glucose supplies most of the energy (Burd et al., 1975; Jones et al., 1975; Pégorier et al., 1977). The plasma lactate/pyruvate ratio is close to 10 (with 100 μ M pyruvate), and higher lactate and (or) lactate/pyruvate ratios are only observed in dystonia, subarachnoid hemorrhage, brain traumas, epilepsies, pyruvate dehydrogenase mutations, and other severe pathological conditions (Owen et al., 1967; Medina, 1985; Fernandez et al., 1986; Mintun et al., 2004; Bjerring et al., 2008; Brody et al., 2008; Rex et al., 2009). In keeping with this, we found pyruvate levels $\sim 120 \mu$ M that are identical to human levels and 40-fold lower than the concentrations used by Zilberter and colleagues. At physiological levels, pyruvate and lactate had no effects on excitatory actions of GABA, DF_{GABA} , and GDPs.

The reduction of DF_{GABA} and GDPs by exceedingly high concentrations of pyruvate (5 mM) is most likely attributable to the acidosis produced at these concentrations. Agents that alter tissue pH alter intracellular chloride (Kaila and Voipio, 1987; Kaila et al., 1993; Chesler, 2003; Glykys et al., 2009; Kim and Trussell, 2009), neuronal excitability, and seizures (Roos and Boron, 1981; Aram and Lodge, 1987; Balestrino and Somjen, 1988; Jarolimek et al., 1989; Kaila, 1994; Bonnet et al., 2000; Dulla et al., 2005, 2009; Ziemann et al., 2008). Propionate, D- and L-lactate reduce GDPs, although only L-lactate is metabolically active (Roos and Boron, 1981; Dulla et al., 2005; Ruussuvori et al., 2010). Therefore, depolarizing actions of GABA are not attributable to the absence of DL-BHB or lactate/pyruvate in the ACSF.

Ketone bodies and GABA signaling

Ketosis (Nehlig and Pereira de Vasconcelos, 1993), like transporters that import D-BHB (Pellerin et al., 1998; Magistretti et al., 1999; Bergersen et al., 2002; Pierre et al., 2002; Rafiki et al., 2003; Vannucci and Simpson, 2003; Erecinska et al., 2004) or BHB

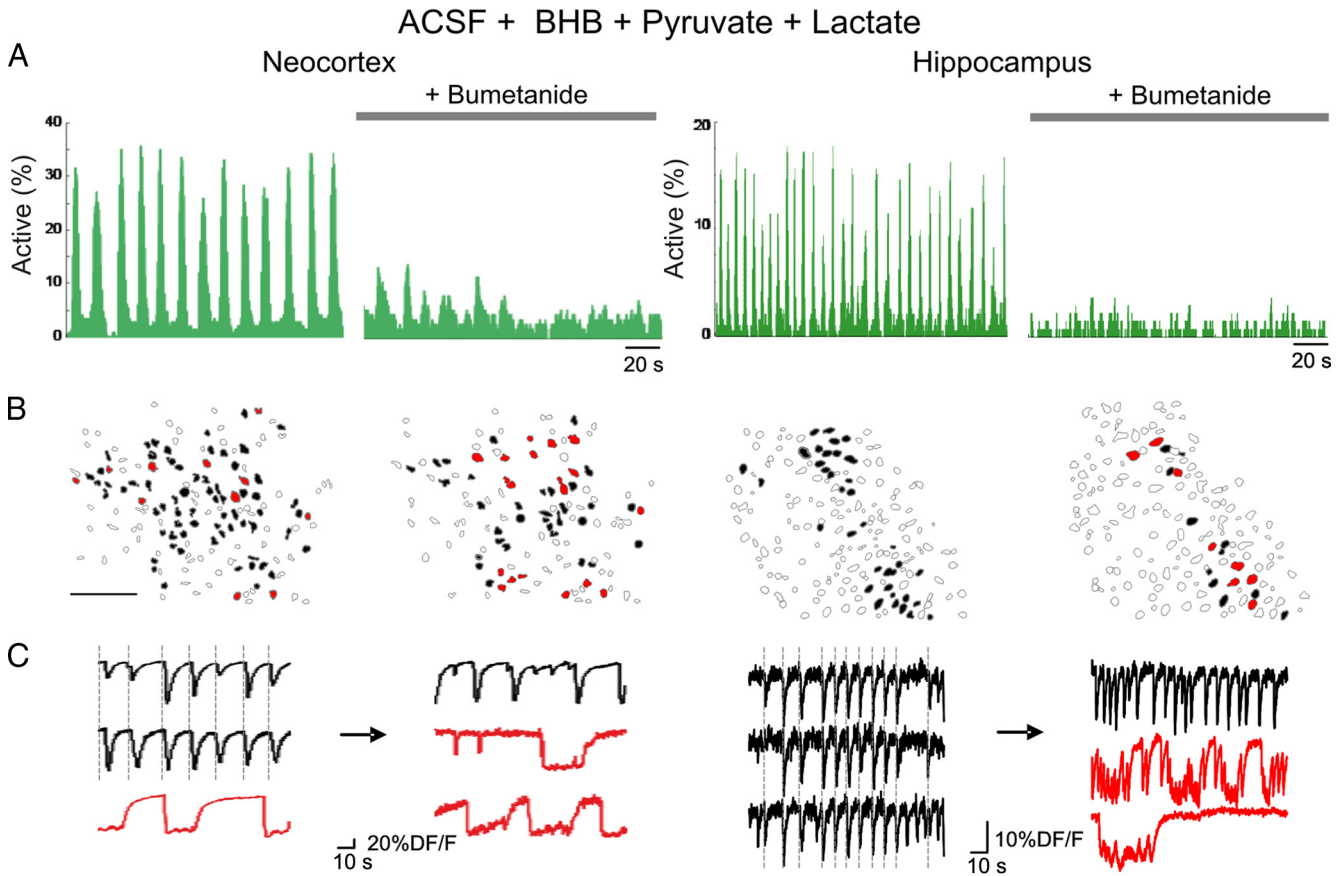


Figure 8. GDPs recorded in the presence of physiological concentrations of lactate, pyruvate, and DL-BHB are similar to those recorded in regular ACSF and are reduced by bumetanide. **A**, Histograms indicating the fraction of active cells as a function of time in calcium movies in slices incubated with 1.5 mM lactate, 150 μ M pyruvate, 5 mM glucose, and 4 mM DL-BHB (Sigma-Aldrich) for at least 1 h. Each peak of synchronous neuronal activity in the histograms corresponds to a GDP. GDPs were strongly reduced in the presence of the NKCC1 antagonist bumetanide (10 μ M) in neocortical and hippocampal slices from P7 rats. **B**, Automatically detected contours of the imaged cells: open contours indicate silent cells, black filled contours indicate cells involved in GDPs, and red filled contours are SPA cells. Note that the number of SPA cells relative to the number of active cells increased in the presence of 10 μ M bumetanide in both the neocortex and hippocampus (scale bar, 100 μ m). **C**, Calcium fluorescence traces of representative cells implicated in GDPs (black) and SPAs (red). Note that some GDP cells display an SPA pattern of activity after adding bumetanide (middle traces).

Table 4. Dynamics of cortical GDPs and SPAs in the neocortex and the hippocampus in the presence of physiological concentrations of lactate, pyruvate, and DL-BHB

ACSF and DL-BHB + Pyruvate + Lactate	Neocortex	Hippocampus
<i>n</i>	5	3
GDP _{freq} (Hz)	0.1 $\pm$ 0.02	0.07 $\pm$ 0.03
GDP _{amp} (%)	60 $\pm$ 10	13 $\pm$ 1
SPA cells (%)	16 $\pm$ 4	3 $\pm$ 0.03

GDP_{freq}, Frequency of occurrence of GDPs (in hertz); GDP_{amp}, fraction of active cells involved in GDPs (percentage); SPA cells, fraction of active cells involved in SPAs (percentage).

dehydrogenase that metabolize it (Page et al., 1971; De Vivo et al., 1975; Leong and Clark, 1984; Bilger and Nehlig, 1991; Clark et al., 1993), peak during development well after the GABA shift. Fatty acid oxidation supports gluconeogenesis (Pégorier et al., 1977), and ketosis acts to “spare glucose for the emergence of audition, vision and more integrated behavior whose appearance during brain maturation seems to critically relate upon active glucose supply” (Nehlig, 2004). GABA currents are not affected by DL-BHB and ketogenic diet reduces seizures generated by GABA receptor antagonists (Appleton and De Vivo, 1973, 1974; Bough and Eagles, 1999; Bough et al., 2000; Thio et al., 2000; Sullivan et al., 2003; Hartman et al., 2007; Yellen, 2008; Maalouf et al., 2009), suggesting, contrary to Zilberter and colleagues, that the antiepileptic actions of ketone bodies are not mediated by GABA signal-

ing. GABA depolarizes immature neurons in nonmammalian animal species and *in utero* in rodents, suggesting that maternal milk and ketone bodies are not required for that effect (Akerman and Cline, 2006; Ben-Ari et al., 2007).

The NKCC1/KCC2 sequence in brain maturation

Contrary to the suggestions of Zilberter and colleagues (Rheims et al., 2009; Holmgren et al., 2010), extensive pharmacological, anatomical, and genetic observations suggest that the removal of chloride in neonatal pups heavily depends on KCC2 (Li et al., 2002; Rheims et al., 2008; Riekkki et al., 2008; Zhu et al., 2008; Takayama and Inoue, 2010). The parallel alterations of KCC2 and GABA polarity have been confirmed in a large variety of animal species from invertebrates to humans (Rivera et al., 1999, 2005; Delpire, 2000; Payne et al., 2003; Sernagor et al., 2003; Dzhalal et al., 2005; Akerman and Cline, 2006; Liu et al., 2006; Ben-Ari et al., 2007; Howard et al., 2007; Kahle et al., 2008; Reynolds et al., 2008; Blaesse et al., 2009; Glykys et al., 2009; Stil et al., 2009; Tanis et al., 2009; Boulenguez et al., 2010). KCC2 and E_{GABA} developmental sequences are unlikely to depend on global metabolic diets because they are cell and sex specific (Kandler and Friauf, 1995; Kandler et al., 2002; Balakrishnan et al., 2003; Gulacsi et al., 2003; Lee et al., 2005; Lohrke et al., 2005; Banke and McBain, 2006; Blaesse et al., 2006; Kim and Trussell, 2009; Belenky et al., 2010). Neurons in which GABA remains depolar-

izing do not express KCC2 (Price et al., 2005; Gilbert et al., 2007; Pozas et al., 2008), and early overexpression of KCC2 in zebra fish embryos (Reynolds et al., 2008) or cortical neurons (Chudotvorova et al., 2005; Lee et al., 2005; Ben-Ari et al., 2007; Cancedda et al., 2007; Wang and Kriegstein, 2008) alter GABA polarity, GABA synapse formation, and neuronal development *in vivo*. KCC2 is downregulated by activity and E_{GABA} shifts accordingly (Woodin et al., 2003; Fiumelli et al., 2005).

Neonatal slices are not energy deprived in 10 mM glucose because they have a low rate of oxygen and glucose consumption and are less susceptible to energy deprivation than adult slices (Cherubini et al., 1989; Novotny et al., 2001; Tyzio et al., 2006). Lowering glucose reduces GDPs, ATP levels, and mitochondrial pH, confirming the sensitivity of GDPs to energy deprivation, but these are not restored by lactate (Takata and Okada, 1995; Wada et al., 1998; Takata et al., 2001) (Ruussuvori et al., unpublished report). Slices and intact hippocampi sustain GDPs for hours (Ben-Ari et al., 1989; Cherubini et al., 1989; Khalilov et al., 1997, 2003; Leinekugel et al., 1997; Safiulina et al., 2006) and are replaced by glutamate-driven early network oscillations when energy is deprived (Allène et al., 2008).

We conclude that the depolarizing action of GABA and related network-driven GDPs in immature cortical slices are not attributable to metabolic insufficiency. Therefore, conventional glucose containing ACSF provides adequate energy supply for cortical slices *in vitro*.

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